



**HAL**  
open science

## **Inulin supplementation lowered the metabolic defects of prolonged exposure to chlorpyrifos from gestation to young adult stage in offspring rats**

Julie Reygner, Lydia Lichtenberger, Ghada Elmhiri, Samir Dou, Narges Bahi-Jaber, Larbi Rhazi, Flore Depeint, Véronique Bach, Hafida Khorsi-Cauet, Latifa Abdennebi-Najar

### ► To cite this version:

Julie Reygner, Lydia Lichtenberger, Ghada Elmhiri, Samir Dou, Narges Bahi-Jaber, et al.. Inulin supplementation lowered the metabolic defects of prolonged exposure to chlorpyrifos from gestation to young adult stage in offspring rats. PLoS ONE, 2016, 11 (10), 10.1371/journal.pone.0164614 . hal-03543026

**HAL Id: hal-03543026**

**<https://u-picardie.hal.science/hal-03543026v1>**

Submitted on 11 Dec 2023

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

RESEARCH ARTICLE

# Inulin Supplementation Lowered the Metabolic Defects of Prolonged Exposure to Chlorpyrifos from Gestation to Young Adult Stage in Offspring Rats

Julie Reygner<sup>1,2</sup>, Lydia Lichtenberger<sup>2</sup>, Ghada Elmhiri<sup>1</sup>, Samir Dou<sup>1</sup>, Narges Bahi-Jaber<sup>1</sup>, Larbi Rhazi<sup>1</sup>, Flore Depeint<sup>1</sup>, Veronique Bach<sup>2</sup>, Hafida Khorsi-Cauet<sup>2</sup>, Latifa Abdennebi-Najar<sup>1</sup>✉\*

**1** UP-EGEAL 2012.10.101, Institut Polytechnique LaSalle Beauvais, Beauvais, France, **2** Laboratoire Périnatalité et Risques Toxiques (PERITOX), UMR-I01 INERIS, Jules Verne University of Picardy, Amiens, France

✉ These authors contributed equally to this work.

\* [latifa.najar60@gmail.com](mailto:latifa.najar60@gmail.com)



CrossMark  
click for updates

## OPEN ACCESS

**Citation:** Reygner J, Lichtenberger L, Elmhiri G, Dou S, Bahi-Jaber N, Rhazi L, et al. (2016) Inulin Supplementation Lowered the Metabolic Defects of Prolonged Exposure to Chlorpyrifos from Gestation to Young Adult Stage in Offspring Rats. PLoS ONE 11(10): e0164614. doi:10.1371/journal.pone.0164614

**Editor:** Hauke Smidt, Wageningen University, NETHERLANDS

**Received:** February 2, 2016

**Accepted:** September 28, 2016

**Published:** October 19, 2016

**Copyright:** © 2016 Reygner et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Data Availability Statement:** All relevant data are within the paper and its supporting information files.

**Funding:** This work was financial supported by the Picardy Regional Council. The funders had no role in study design, data collection and analysis, decision to publish or preparation of the manuscript.

**Competing Interests:** The authors have declared that no competing interests exist.

## Abstract

Increasing evidence indicates that chlorpyrifos (CPF), an organophosphorus insecticide, is involved in metabolic disorders. We assess the hypothesis whether supplementation with prebiotics from gestation to adulthood, through a modulation of microbiota composition and fermentative activity, alleviates CPF induced metabolic disorders of 60 days old offspring. 5 groups of Wistar rats, from gestation until weaning, received two doses of CPF pesticide: 1 mg/kg/day (CPF1) or 3.5 mg/kg/day (CPF3.5) with free access to inulin (10g/L in drinking water). Then male pups received the same treatment as dams. Metabolic profile, leptin sensitivity, insulin receptor (IR) expression in liver, gut microbiota composition and short chain fatty acid composition (SCFAs) in the colon, were analyzed at postnatal day 60 in the offspring (PND 60). CPF3.5 increased offspring's birth body weight (BW) but decreased BW at PND60. Inulin supplementation restored the BW at PND 60 to control levels. Hyperinsulinemia and decrease in insulin receptor  $\beta$  in liver were seen in CPF1 exposed rats. In contrast, hyperglycemia and decrease in insulin level were found in CPF3.5 rats. Inulin restored the levels of some metabolic parameters in CPF groups to ranges comparable with the controls. The total bacterial population, short chain fatty acid (SCFA) production and butyrate levels were enhanced in CPF groups receiving inulin. Our data indicate that developmental exposure to CPF interferes with metabolism with dose related effects evident at adulthood. By modulating microbiota population and fermentative activity, inulin corrected adult metabolic disorders of rats exposed to CPF during development. Prebiotics supply may be thus considered as a novel nutritional strategy to counteract insulin resistance and diabetes induced by a continuous pesticide exposure.

## Introduction

During the last decade, the incidence of obesity and diabetes has dramatically increased all over the world. According to the World Health Organization (WHO), the world prevalence of diabetes estimated to be at 2.8% in 2000 will reach 4.4% in 2030 [1]. Western diet and lack of physical exercise are currently related to this burden of metabolic diseases. Recently, epidemiological and animal studies pointed out the involvement of chemical exposure during pregnancy and lactation in the increasing incidence of this metabolic syndrome [2–5].

Although it is increasingly restricted to the US and Europe, the highly-lipophilic organophosphorus (OP) compound Chlorpyrifos (CPF) is one of the most frequently used non-persistent insecticides worldwide and is commonly found in fruits and vegetables [6]. Numerous studies at high level exposure have endorsed the neurotoxic effects of CPF in both human and animal models [7]. CPF exerts its systemic toxicity by irreversibly inhibiting acetylcholinesterase [8]. At low-levels this compound targets cell signaling cascades that govern neuronal and hormonal signals, which are linked to homeostatic balance and cellular differentiation.

Recent epidemiological studies and investigations in experimental animal models support the effect of early exposure of CPF in the ontogeny of diabetes [6,9,10]. Neonatal exposure to CPF displays hyperinsulinemia and hyperlipidemia in adulthood rat, two major risk factors for type 2 diabetes mellitus (T2D) and atherosclerosis. These findings extend Barker's hypothesis [11] showing that in the absence of intrauterine growth restriction, CPF during early development can result in permanent changes in the physiology and metabolism resulting in increased metabolic risks in adulthood.

Gut microbiota exerts a significant role in the pathogenesis of the metabolic syndrome as confirmed by studies conducted both in human and animal models [12–14]. In fact, gut microbiota plays a great variety of functional roles impacting human physiology. It modulates host nutrition by the production of vitamins and fermentation of food components indigestible by the host, protects against pathogens [15] and drug metabolism and influences intestinal epithelial homeostasis [16]. An impairment of the fine balance between gut microbes and host's interactions induces the intestinal translocation of bacterial fragments and the development of "metabolic endotoxemia", leading to systemic inflammation and insulin resistance [17]. Clinical and animal studies on obesity and T2D showed a shift in the pattern of the gut microbiota, in particular a decrease in the ratio of *Firmicutes/Bacteroidetes* for obesity [14] and a lower proportion of Clostridiales for T2D [18]. Compelling evidence suggests that oral supplementation with selectively fermented oligosaccharides (known as prebiotics) improves these metabolic disorders via several mechanisms [19–21]. Moreover, prebiotics are likely associated with the increase in Bifidobacteria and Lactobacilli and the production of short chain fatty acids (SCFAs), which are involved in the modulation of the host metabolism [22]. For example, feeding genetically or diet-induced obese mice with prebiotics significantly increased the abundance of *Akkermansia muciniphila*, which was correlated with an improved metabolic status [21,23]. Other studies have shown that prebiotics reinforce the gut barrier, increase satiety by promoting gut hormones, improve glucose tolerance, counteract hepatic steatosis (lipogenesis) and insulin resistance [24].

We recently showed that neonatal exposure to CPF disturbed the microbiota composition specifically the proportion of Lactobacilli in the colon at PND 60 [25]. In the current study, we wanted to test whether early prebiotic supplementation counteracts the metabolic disorders induced by early exposure to CPF. Rats were exposed from pregnancy to weaning, to two doses of CPF (1mg/Kg/day, CPF1) and (3.5 mg/Kg/day, CPF3.5), either alone or in association with inulin. Inulin was administered along with drinking water at a dose of 10g/L. We intended to induce CPF exposure from gestation to weaning to mimic the effect of CPF exposure in a real

human neonate during its development. We evaluated the effect of the different treatments in rats on lipid and glucose metabolism, insulin and leptin, gut microbiota composition and SCFA production at 60 days of age. The supplementation with inulin is relevant in this study because it may help to identify new properties of prebiotics in both the mother and the child exposed to pesticides, and to develop new strategies against metabolic programming in later life.

## Materials and Methods

### Chemicals

Chlorpyrifos (O, O-diethyl, O-(3,5,6-trichloro-2-pyridyl) phosphorothioate), purity  $99.8\% \pm 0.1\%$ , was supplied by LGC Standards (Molsheim, France). It was dissolved in rapeseed oil (MP Biomedicals, Illkirch, France), which served as a vehicle and administered daily by gavage at a dose of 1 mg/kg of BW/day (CPF1) or 3.5 mg/kg of BW/day (CPF3.5). A commercially available product of chicory inulin, with a dry matter of 96%, containing 90% inulin with an average polymerization degree of 10% and a free sugar content of 10% was added to drinking water and the average inulin consumption was  $3.73 \text{ mg} \pm 0.04 \text{ mg/g}$  of BW/day.

### Animals and treatment

**Dams.** All procedures were carried out according to the Animal Care and Use Committee at Jules Verne University of Picardy (n°291112–19, Amiens, France) which approved this study. All efforts were made to minimize animal's suffering. Wistar rats, thirty-two females and five males (Janvier, Le Genest St Isle, France) were housed in breeding cages under constant conditions of ambient temperature (23°C), hygrometry (26%), with a 12h light/dark cycle and free access to food and water. After 1 week of acclimation period, females were mated with males (2 females per male). Time-pregnant, primiparous Wistar rats were determined by the presence of spermatozoa in vaginal smear. Pregnant females were individually housed and randomly assigned (1:1) to five treatment groups ( $n = 5$  to 6) and a control group ( $n = 5$ ). In each treatment group, the dams, from the first gestation day (GD) until lactation day (LD) 21, were exposed to a daily gavage of vehicle or CPF associated with or without inulin. The different treatment groups were as follows: CPF groups were CPF0, CPF1 and CPF3.5 and inulin groups were inu0 and inu1. CPF amounts were adjusted daily according to any changes in the body weight and administered at approximately the same time each morning. Each pregnant female's food and drink intake were recorded every three days from GD 0 to LD 21.

**Pups.** At postnatal day 1 (PND 1), all pups were counted, sexed and weighed. Each litter was homogenized and adjusted to 8 pups. At PND 21, only male pups were weaned and received the same treatment as dams. Birth weight, food and drink intake were recorded every 2 days from PND 1 to PND 60. At PND 60, animals were euthanized by intraperitoneal administration of lethal dose of sodium pentobarbital (Ceva Santé Animale (Libourne, France) and a sample of blood, brain, liver, fecal content and colon were collected. Tissues were immediately frozen and stored at  $-80^\circ\text{C}$ . Blood samples were centrifuged at 4000g for 10min and plasma was collected, aliquoted and stored at  $-80^\circ\text{C}$ .

### Metabolic assay

Plasma leptin and insulin levels were measured by specific commercial RIA kit (EMD, Millipore, France). Standard spectrophotometric methods based on an automation program by Amiens University Hospital were used for measurement of the following serum parameters:

Cholesterol, High Density Lipoprotein (HDL), Low Density Lipoprotein (LDL), triglycerides (TG), glycaemia, alanine transaminase (ALT) and aspartate transaminase (AST).

### Protein extraction from liver

Proteins were extracted from 20 mg of liver tissue according to methods described in our previous studies [26,27]. Briefly, tissues were lysed in 1X RIPA Buffer containing a cocktail of protease inhibitors (Thermo Scientific, France). Lysis was performed using a Tissue Lyser device (Qiagen, France). Samples were incubated for 15–20 min at 4°C and centrifuged (14000g; 15 min; 4°C). The supernatant was removed and stored at -80°C. Protein concentrations were measured using a Pierce BCA Protein Assay kit according to the manufacturer's instructions.

### Gel electrophoresis and Western Blot analysis

For gel electrophoresis, the protein samples were resuspended and heated for 5 min at 95°C and loaded on a 4–12% SDS-PAGE (Criterion XT Bis-Tris Gel, Biorad, France). Proteins were transferred onto a nitrocellulose membrane (Biorad, France) after electrophoresis. Membranes were blocked with TBS-T/5% milk and incubated overnight at 4°C with the primary antibody anti insulin receptor  $\beta$  (IR $\beta$ ) (rabbit monoclonal, #3020, Cell Signaling, diluted 1/1000). The blot was then incubated with peroxidase conjugated secondary antibody (Abcam, diluted 1/5000). The protein signal was detected using the ECL kit (Amersham Biosciences). Proteins were analyzed using anti- Signals on autoradiographic films were quantified by scanning densitometry using ImagQuant 350 (GE Healthcare, France).

### DNA Extraction and 16S RNA qPCR analysis

Total bacterial count and specific bacterial profile were evaluated by quantitative PCR analyses targeting bacterial group-specific 16S rRNA genes using the Rotor-Gene system (Qiagen, France). Total DNA was extracted from 20 to 25 mg of colon content using the Qiagen QIAamp Fast DNA stool kit according to the manufacturer's instructions (Qiagen, France). PCR inhibitions were tested with TaqMan<sup>®</sup> Exogenous Internal Positive Control and the TaqMan<sup>®</sup> universal master Mix (Life Technologies S.A, France). No PCR inhibition was detected using 10<sup>-3</sup> dilutions for the feces samples. DNA from each sample was amplified using selected primers and probe sets given in Table 1 [28,29]. For bacterial SYBR-green amplification studies, a melting curve was added to show the amplification specificity and the following PCR profile was used: 1 cycle at 95°C for 12 min, followed by 40 cycles of 95°C for 15 s, 60°C for 30 s, 72°C for 30 s. Cycle amplification data were quantified according to standard curves. Data were expressed as log<sub>10</sub> (copy number)/g of feces.

### Short Chain Fatty Acid (SCFA) analysis

The concentration of acetate, propionate and butyrate in the fecal material were determined after water extraction of acidified samples using gas-liquid chromatography (Thermo Scientific, Focus GC-AutoInjector AI 3000) as described in our previous study [30].

### Statistical analysis

Statistical analyses were performed with StatView software (version 5.0, Abacus Concepts Inc., Berkeley, CA, USA). The non-parametric tests Kruskal Wallis was used to analyze the effects of CPF ([CPF0, CPF1, CPF3.5] inu0, [CPF0, CPF1, CPF3.5] inu1) followed by Mann Whitney test, when significance reached  $p < 0.05$ . Effects of inulin was analyzed by Mann Whitney test ([inu0, inu1] CPF0, [inu0, inu1] CPF1, [inu0, inu1] CPF3.5). All results were presented as

**Table 1. Primers and probes used in this study.**

	Target	Primers and Probes	Sequences (5'-3')
TaqMan System	All bacteria <sup>a</sup>	F-Bact 1369	CGGTGAATACGTTCCCGG
		R-Prok 1492	TACGGCTACCTTGTTACGACTT
		<b>P-TM1389F</b>	<b>6FAMCTTGTTACACACCGCCCGTC</b>
	<i>Bifidobacterium</i> <sup>a</sup>	F-Bifid 09c	CGGGTGAAGTAATGCGTGACC
		R-Bifid 06	TGATAGGACGCGACCCCA
		<b>P-Bifid</b>	<b>6 FAM CTCTGGAACGGGTG</b>
	<i>Clostridium leptum</i> group <sup>a</sup>	F-Clept09	CCTTCCTGCGSAGTTA
		R-Clept 08	GAATTAACCACATACTCCACTGCTT
		<b>P-Clept 01</b>	<b>6 FAM-CACAATAAGTAATCCACC</b>
<i>Clostridium coccooides</i> group <sup>a</sup>	F-Ccoc07	GACGCCGCGTGAAGGA	
	R-Ccoc14	AGCCCCAGCCTTTCACATC	
	<b>P-Erec482</b>	<b>VIC-CGGTACCTGACTAAGAAG</b>	
SyberGreen System	<i>Bacteroidetes</i> <sup>b</sup>	BactF	CCTWCGATGGATAGGGGTT
		BactR	TCCCCAGGTGGAATACTTAACG
	<i>Lactobacillus/Leuconostoc/Pediococcus</i> <sup>a</sup>	F-lacto05	AGCAGTAGGGAACTCTCC
		R-lacto04	CGCCACTGGTGTTCYTCATATA
	<i>Firmicutes</i> <sup>b</sup>	FirmF	ACCCGCGTCTGATTAGCTAGTT
		FirmR	CCTCTCAGGCCGGCTACTG

<sup>a</sup> [28]

<sup>b</sup> [29]

doi:10.1371/journal.pone.0164614.t001

means ± standard error of the mean (SEM). Significance was set at the value of  $p < 0.05$  and the indicative results were presented if relevant. Due to birth body weight variation among the different groups, body weight change from birth to PND60 was adjusted accordingly to birth body weight (BW PND1):  $((\text{PND60-BW PND1})/\text{BW PND1})$ .

## Results

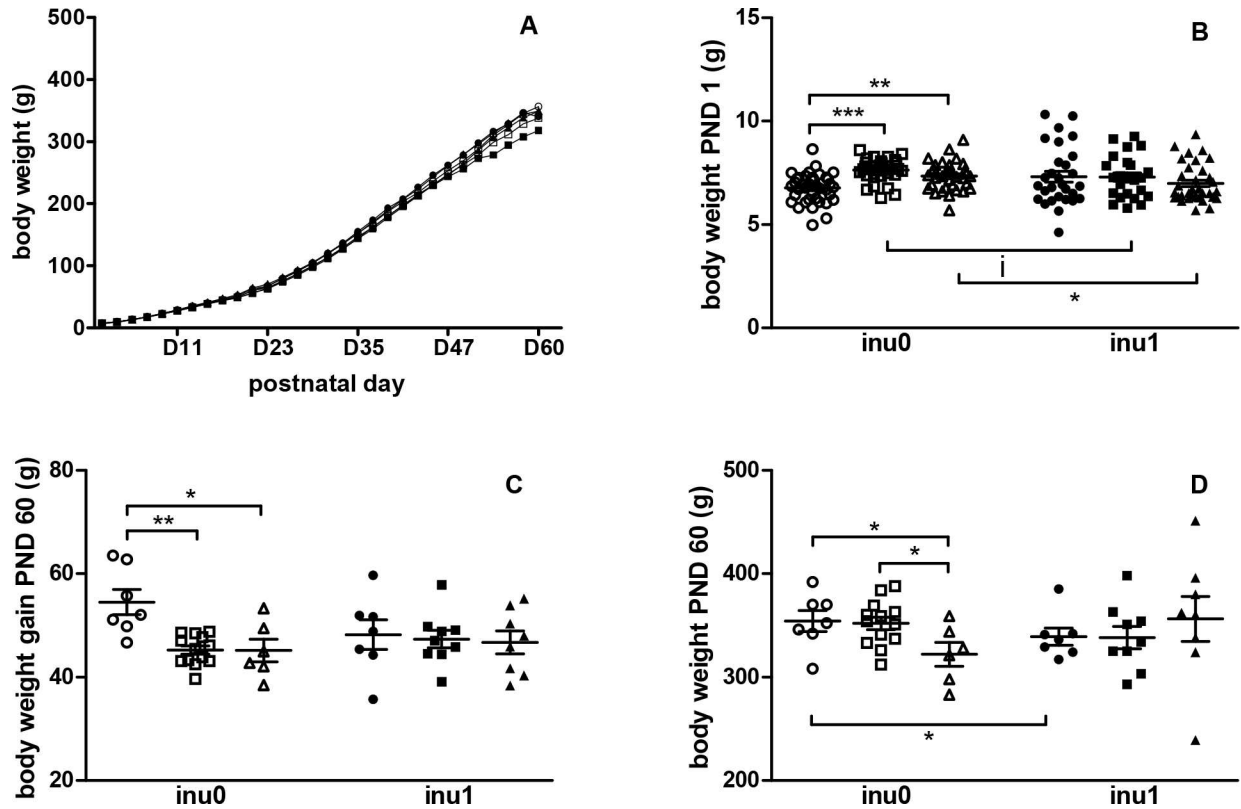
### Dam's body weight, food and drink intake (GD 0 and LD 21)

Dam's body weight, food intake and drink intake were measured every 3 days. Neither CPF nor inulin significantly affected maternal weight gain, food intake and drink intake (S1 Table). No signs of cholinergic toxicity such as tremor, salivation or diarrhea were observed in animals during the whole experiment.

### Offspring's body weight and growth, food and drink intake

Offspring's body weight, growth and food intake were recorder every two days from PND1 to PND60. The global growth curve of different groups did not appear to be affected by treatments except CPF3.5 animals. Indeed, a "drop" of the curve for these exposed rats to the highest dose of CPF was observed at PND53 (Fig 1A). The mean BW of control offspring group at birth and PND 60 were  $6.77 \pm 0.12$  g and  $355.9 \pm 8.2$  g, respectively. Inulin alone did not affect offspring BW and BW gain but animal's weight at PND 60 decreased with inulin supplementation ( $p < 0.05$ ) (Fig 1D). Gestational exposure to CPF significantly increased BW at birth ( $p < 0.001$ ) (Fig 1B). BW gain and BW at PND 60 were decreased in both CPF-exposed groups ( $p < 0.05$ ) (Fig 1C and 1D). This reduction in weight gain was not associated with changes in either food or water intake (S2 Table). The supplementation of inulin in CPF-exposed groups induced a decrease of BW at birth as compared with CPF animals. However, no effects of inulin





**Fig 1. Offspring's body weight and growth.** (A) Global growth of animals from PND1 to PND 60; (B) Body Weight at PND 1 [CPF0inu0 n = 38/CPF1inu0 n = 31/CPF3.5 n = 35/CPF0inu1 n = 29/CPF1inu1 n = 25/CPF3.5inu1 n = 34]; (C) BW gain PND 60; (D) BW at PND 60 of male pups. The results are expressed as mean  $\pm$  SEM and individual values and analyzed by Mann Withney test. Control groups (CPF0, circles), CPF-exposed groups (CPF1: 1 mg/kg/day, squares; CPF3.5: CPF3.5 mg/kg/day, triangles) or inulin groups (black symbol). Signification \* $p < 0.05$ ; \*\* $p < 0.01$ ; \*\*\* $p < 0.001$ , i = 0.058.

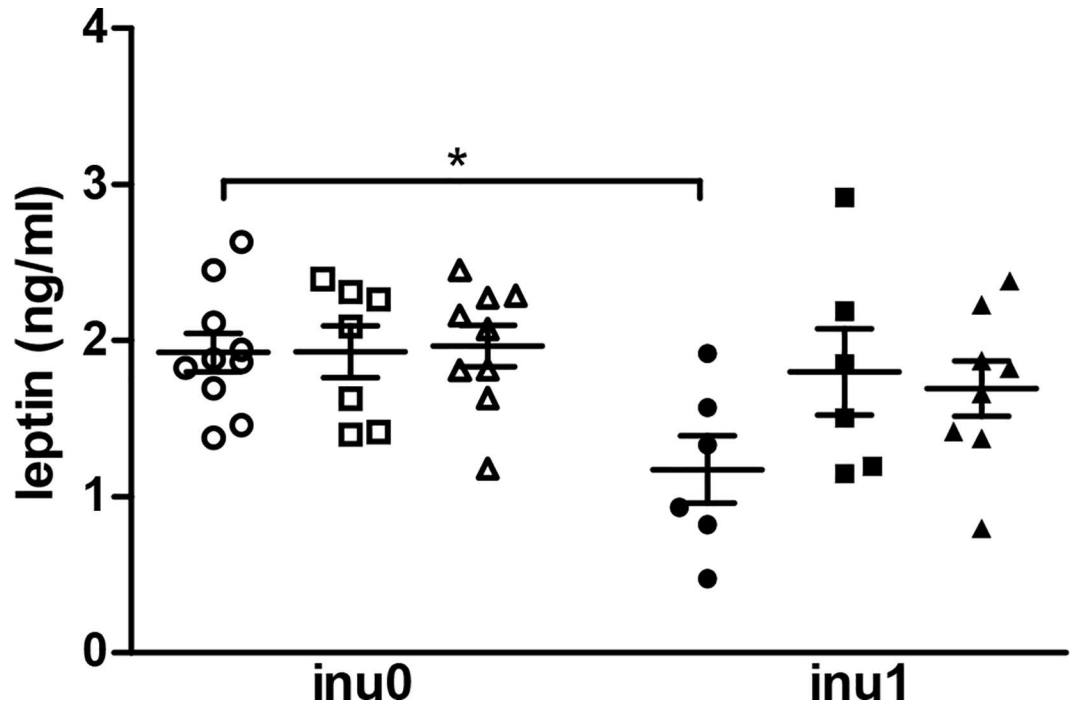
doi:10.1371/journal.pone.0164614.g001

on BW gain and BW at PND 60 were observed in CPF-exposed animals (Fig 1C and 1D). Thus CPF exposure modified the body weight growth pattern of the offspring's from birth to weaning. In both CPF exposed rats, inulin supply induced a decrease of birth body weight and restored the BW at PND 60 to control levels.

### Metabolic parameters

The effect of the different treatments on metabolic parameters was compared between the different groups. Inulin alone did not affect metabolic parameters while leptin level at PND 60 decreased when inulin was supplied in the drinking water ( $p < 0.05$ ) (Fig 2). CPF exposure during pregnancy and lactation induced different effects on the metabolic parameters depending on the dose used: CPF3.5 group showed a significant higher increase in glycaemia at PND 60 ( $p < 0.05$ ) compared to the control (Fig 3A). A slight increase in plasma insulin level ( $p = 0.08$ ) at PND 60 was observed in CPF1-exposed offspring's while a slight decrease in insulin secretion was observed in animals exposed to the higher dose ( $p = 0.08$ ). When supplemented with inulin, the levels of insulin in both CPF1 and CPF3.5 groups were close to the values observed in control animals ( $p < 0.05$ ) (Fig 3B).

The lipid status (i.e., the total cholesterol, LDL and HDL) was not affected by inulin or CPF exposure alone except for the level of TG which was lower in CPF3.5-exposed rats as compared to control animals ( $p < 0.05$ ). The TG level was recovered when animals were co-exposed with



**Fig 2. Leptin level of male pups at PND 60.** Data are expressed as mean  $\pm$  SEM [CPF0inu0 n = 10/CPF1inu0 n = 7/CPF3.5 n = 9/CPF0inu1 n = 6/CPF1inu1 n = 6/CPF3.5inu1 n = 8]. Control groups (CPF0, circles), CPF-exposed groups (CPF1: 1 mg/kg/day, squares; CPF3.5: CPF3.5 mg/kg/day, triangles) or inulin groups (black symbol). Signification \* $p < 0.05$ ; \*\* $p < 0.01$ , \*\*\* $p < 0.001$ .

doi:10.1371/journal.pone.0164614.g002

inulin and CPF3.5 in comparison to CPF3.5 animals alone ( $p < 0.01$ ) (Fig 4). Fig 5A shows an increase in ALT enzyme for the CPF3.5-exposed groups ( $p < 0.05$ ) and a slight increase ( $p = 0.07$ ) in ALT in CPF1 exposed animals compared to the control. The difference in ALT level did not persist when animals received both CPF3.5 and inulin. Meanwhile, no changes in AST level were observed with the different treatments (Fig 5B). Thus CPF modified the metabolic status of the offspring's and inulin restored the levels of some metabolic parameters in CPF groups to ranges comparable with the controls.

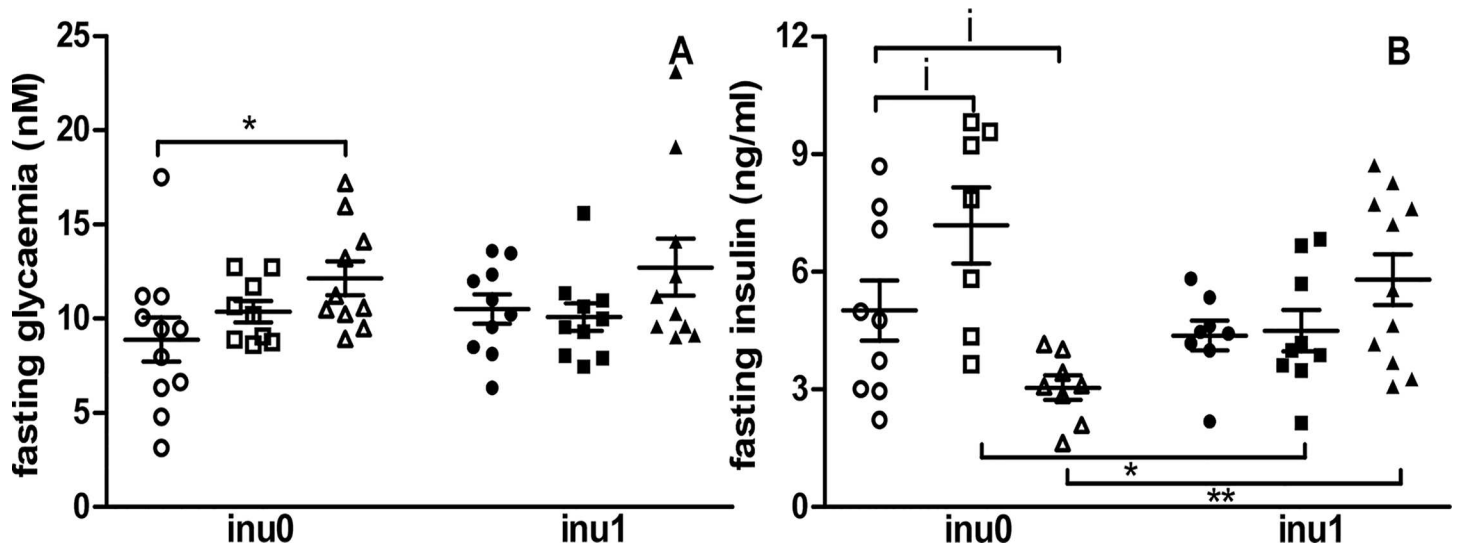
### Insulin protein expression

Western blot analysis evidenced a significant ( $p < 0.05$ ) decrease of IR $\beta$  protein expression in the liver of CPF1 as compared to controls. When supplemented with inulin, IR $\beta$  was significantly ( $p < 0.01$ ) recovered only in CPF1-exposed group (Fig 6A and 6B).

### Gut microbiota

Total bacterial count and specific bacterial profile were evaluated by quantitative PCR analyses targeting bacterial group-specific 16S rRNA. Perinatal exposure to inulin alone significantly increased the global bacterial population ( $p < 0.05$ ) (Fig 7A) and total SCFA production ( $p < 0.01$ ) with a significant increase in the proportion of butyric acid ( $p < 0.05$ ) (Table 2). A significant decrease in the population of *Firmicutes* (Fig 7B) was observed in CPF-exposed animals ( $p < 0.05$ ) for CPF1 and a slight decrease ( $p = 0.08$ ) for CPF3.5 respectively. Perinatal exposure to CPF alone significantly decreased *C. coccoides* group ( $p < 0.05$ ) and tended to reduce *C. leptum* count too (Fig 8A and 8B). In the CPF3.5+inu group, *C. coccoides* group was





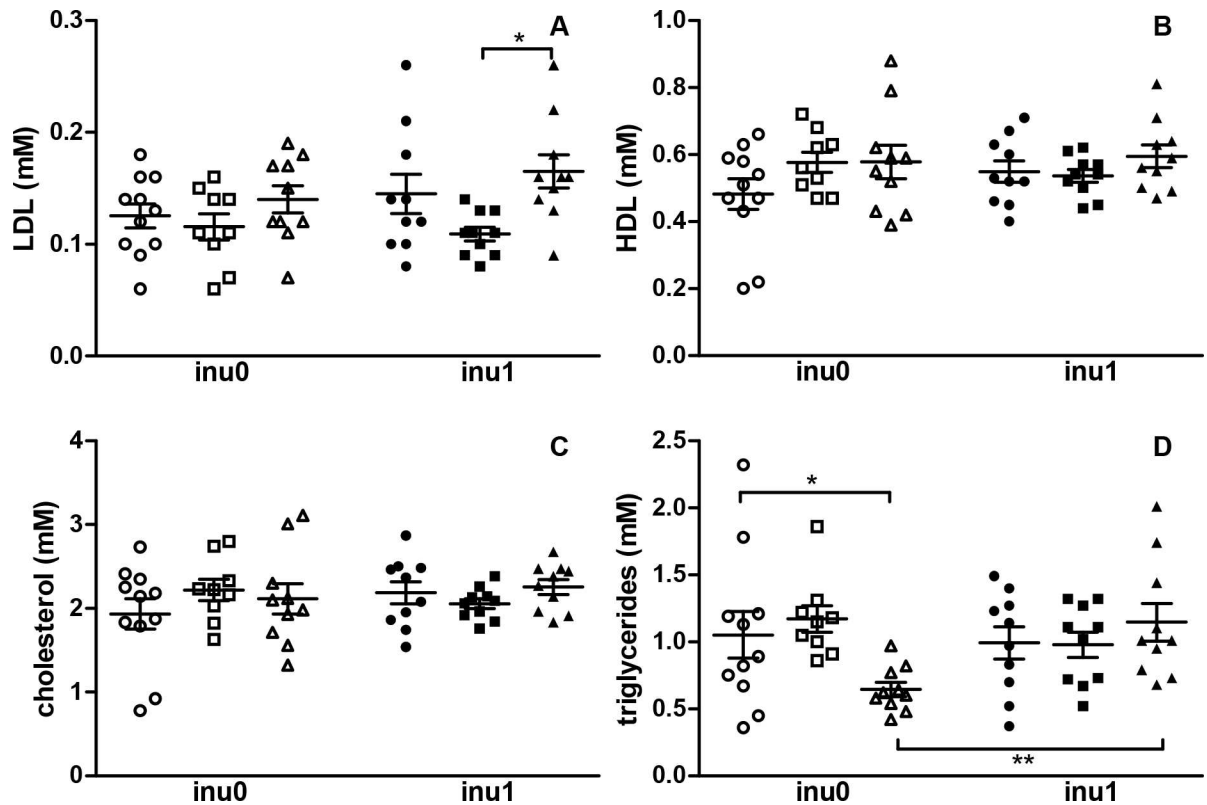
**Fig 3. Fasting Glycaemia and Insulin profile.** (A) Fasting Glycaemia [CPF0inu0 n = 11/CPF1inu0 n = 9/CPF3.5 n = 10/CPF0inu1 n = 10/CPF1inu1 n = 10/CPF3.5inu1 n = 10]; (B) Fasting Insulin are expressed as mean  $\pm$  SEM [CPF0inu0 n = 9/CPF1inu0 n = 7/CPF3.5 n = 8/CPF0inu1 n = 8/CPF1inu1 n = 9/CPF3.5inu1 n = 11]. Control groups (CPF0, circles), CPF-exposed groups (CPF1: 1 mg/kg/day, squares; CPF3.5: CPF3.5 mg/kg/day, triangles) or inulin groups (black symbol). Signification \* $p < 0.05$ ; \*\* $p < 0.01$ ; \*\*\* $p < 0.001$ , i = 0.08.

doi:10.1371/journal.pone.0164614.g003

significantly ( $p < 0.01$ ) higher than CPF3.5-exposed rats (Fig 8A). The *Firmicutes/Bacteroidetes* ratio decreased only in the CPF1 group in comparison to the control ( $p < 0.05$ ) (Fig 7D). In the CPF1+inu group, the total count of *Firmicutes* population and *C. leptum* group were still significantly lower when compared to the control group ( $p < 0.05$ ) (Figs 7B and 8B). For the production of SCFAs, supplementation of CPF1 animals with inulin increased the total SCFAs production ( $p < 0.05$ ) as well as the proportion of butyric acid ( $p = 0.058$ ) compared to CPF1 animals without inulin. However, in CPF3.5 group, inulin supplementation only increased the proportion of butyric acid ( $p = 0.058$ ) when compared to CPF3.5 without inulin. Of note, a high decrease of total SCFAs was observed in CPF3.5 animals supplemented with inulin as compared to the group receiving only inulin ( $305 \pm 16 \mu\text{mol/g}$  vs.  $393 \pm 27,1 \mu\text{mol/g}$  respectively,  $p < 0.01$ ). Thus, CPF decreased *Firmicutes* population. Inulin supplementation enhanced the total bacterial population, SCFA production and butyrate levels in CPF groups.

## Discussion

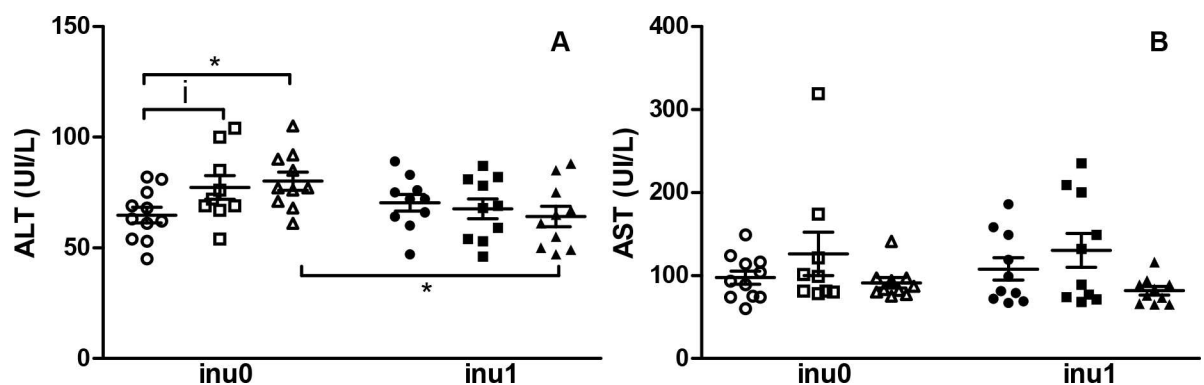
This study demonstrates that CPF exposure during a critical window of development induced permanent defects in normal newborn male rats in terms of their phenotype and their metabolic regulation during adulthood. The exposure to CPF during gestation led to an increase in BW when the offspring were born. However, a long term exposure extending from gestation to adulthood induced a decrease in BW gain and BW in the offspring at day 60. Perturbations in the perinatal environment can impair metabolic programming that can consequently increase susceptibility to T2D in adulthood. Several studies addressing the role of developmental exposure to pesticides with diabetes have rapidly expanded over the past years suggesting that they contribute to metabolic programming [9,31]. In our study we demonstrate that a continuous exposure to a “safe” dose of CPF beginning in the womb until adult life impairs adult metabolism of the offspring’s leading to insulin and lipid dysregulation. Based on these findings, one can assume that the exposure to CPF during gestation, lactation and even during other sensitive periods of development, as pre-puberty, impairs developmental programming. Although



**Fig 4. Plasma lipids status.** (A) LDL; (B) HDL; (C) cholesterol; (D) triglycerides are expressed as mean ± SEM [CPF0inu0 n = 11/CPF1inu0 n = 9/CPF3.5 n = 10/CPF0inu1 n = 10/CPF1inu1 n = 10/CPF3.5inu1 n = 10]. Control groups (CPF0, circles), CPF-exposed groups (CPF1: 1 mg/kg/day, squares; CPF3.5: CPF3.5 mg/kg/day, triangles) or inulin groups (black symbol). Signification \*p<0.05; \*\*p<0.01, \*\*\*p<0.001.

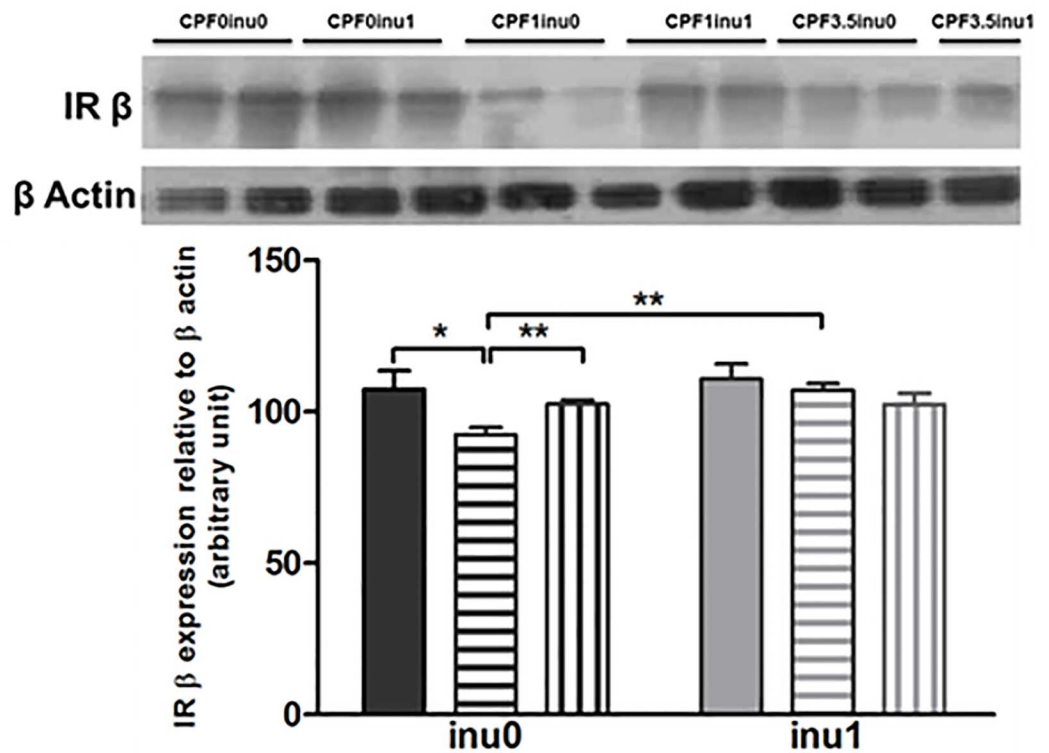
doi:10.1371/journal.pone.0164614.g004

our design does not able determining precisely which of the different targets sensitive windows were involved in the “programmed” metabolic defects, it complements the Barker hypothesis which makes a link between low birth weight and diabetes [32], extending the same outcomes into the situation to developmental exposure to pesticides, even in the absence of growth



**Fig 5. Plasma liver enzymes profile.** (A) ALT; (B) AST are expressed as mean ± SEM [CPF0inu0 n = 11/CPF1inu0 n = 9/CPF3.5 n = 10/CPF0inu1 n = 10/CPF1inu1 n = 10/CPF3.5inu1 n = 10]. Control groups (CPF0, circles), CPF-exposed groups (CPF1: 1 mg/kg/day, squares; CPF3.5: CPF3.5 mg/kg/day, triangles) or inulin groups (black symbol). Signification \*p<0.05; \*\*p<0.01, \*\*\*p<0.001, i = 0.07.

doi:10.1371/journal.pone.0164614.g005

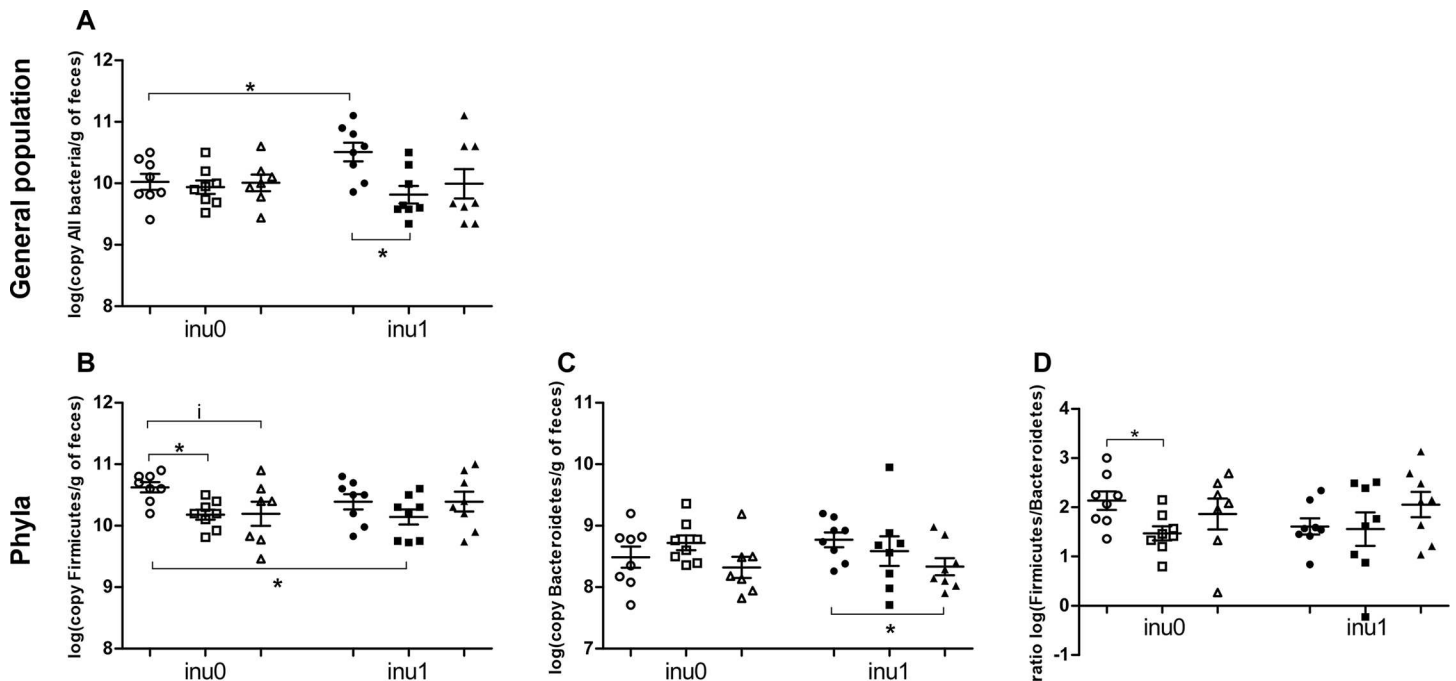


**Fig 6. Determination of IR $\beta$  in liver of rats at PND 60 by Western blot.** Data are expressed as mean  $\pm$  SEM [CPF0inu0 n = 5/CPF1inu0 n = 5/CPF3.5 n = 5/CPF0inu1 n = 5/CPF1inu1 n = 4/CPF3.5inu1 n = 4]. Control groups (CPF0, circles), CPF-exposed groups (CPF1: 1 mg/kg/day, squares; CPF3.5: CPF3.5 mg/kg/day, triangles) or inulin groups (black symbol). (n = 8 to 10). Signification \*p<0.05; \*\*p<0.01,\*\*\*p<0.001.

doi:10.1371/journal.pone.0164614.g006

retardation. The specific pattern of birth weight increase and BW loss after pre- and postnatal exposure to CPF differs from studies published by others on exposure to CPF [33] or Parathion [34], which is another organophosphorous in neonates. This discrepancy can be explained by the different doses used, the time window, the type (acute or chronic) of exposure and the mode of administration (oral, subcutaneous) in different studies.

The major drastic effects of CPF exposure on glucose and lipid metabolism seen in the 60 days old offspring rats were observed in those subjected to the higher dose of 3.5 mg/kg/day CPF. Indeed, an increase in glycaemia concomitant with a drastic reduction of insulin level was observed. This result may reflect an alteration of the pancreas integrity and functionality and as a consequence, the inability of the pancreas to control glycaemia in CPF-exposed rats. Moreover, the alteration of glucose metabolism observed for the higher dose of CPF resulted in a significant increase in serum alanine aminotransferase reflecting hepatic injuries. Indeed, Mansour et al., showed that CPF caused oxidative damage leading to impaired liver cell membrane permeability and thus the release of hepatic enzymes [35]. Furthermore, in association with the decrease in insulin secretion, a subsequent decrease in triglycerides was seen in CPF3.5-exposed rats. As insulin is an important regulatory factor of lipid metabolism [36], one can assume that an alteration in insulin secretion perhaps modified the triglyceride content. Other mechanisms can also contribute to the disturbance of triglyceride synthesis, notably the ones targeting adipose tissue and/or liver signaling that are essential for homeostasis regulation [37,38]. We recently reported that CPF exposure increased para-cellular permeability in the



**Fig 7. General bacterial profile.** (A) All Bacteria; (B) Firmicutes; (C) Bacteroidetes; (D) Firmicutes/Bacteroidetes ratio; are expressed as log(number of copy/g of feces) mean ± SEM [CPF0inu0 n = 8/CPF1inu0 n = 8/CPF3.5 n = 7/CPF0inu1 n = 8/CPF1inu1 n = 8/CPF3.5inu1 n = 8]. Control groups (CPF0, circles), CPF-exposed groups (CPF1: 1 mg/kg/day, squares; CPF3.5: CPF3.5 mg/kg/day, triangles) or inulin groups (black symbol). Signification \*p<0.05; \*\*p<0.01, \*\*\*p<0.001, i = 0.09.

doi:10.1371/journal.pone.0164614.g007

small intestine [39] and delayed intestinal epithelial maturation [25], which may suggest a modification of intestinal absorption of triglycerides in the blood stream.

In agreement with the studies by Slotkin [9,10], CPF1-exposed rats presented a hyperinsulinemia. This result showed that an apparently “safe” exposure was probably maintained in the CPF1-group by the compensatory hypersecretion of insulin. This situation is close to what was observed in a pre-diabetic state. We also reported that the liver of CPF1 animals contain reduced amounts of IRSβ, which support our hypothesis and reflect a compensatory response to chronically elevated insulin level and alteration of insulin sensitivity [40]. These findings highlighted a mechanistic link between insulin signaling in the liver and the subsequent emergence of hyperinsulinemia and hyperlipidemia.

The original approach used in this study was based on the use of inulin, acting as a prebiotic to counteract the side effects of CPF on metabolism. Prebiotics are known to improve the host's health by inducing favorable changes in intestinal microbiota [41]. Inulin selectively stimulates beneficial Bifidobacteria and Lactobacilli *in vitro* [42,43] and also in human subjects and rodents [44–46].

Intestinal SCFA concentration, especially butyrate, the preferred energy source of colonocytes, increases when inulin is consumed [45,46]. In agreement with this, inulin supplementation resulted in an increase in the general population of bacteria and the total SCFA production. An increase in SCFA, observed in supplemented groups, may explain the reduction of insulin level noticed in CPF1-exposed rats because of the role of SCFAs in the activation of G-protein–coupled free fatty acid receptor (GPR43) in the adipose tissue [47]. Moreover, the observed increase in butyrate in rats consuming inulin is probably due to a cross-feeding phenomenon such that the butyrate-producing bacteria belonging to *C. coccooides* (cluster XIVa) and *C. leptum* group (cluster IV) such as *Roseburia intestinalis* or *Faecalibacterium*

**Table 2. Bacteria fermentation activity in offspring at PND 60.**

	CPF0		CPF1		CPF3.5	
	inu0	inu1	inu0	inu1	inu0	inu1
SCFA (μmol/g)	289.5±15.3	393.0±27.1 <sup>**a</sup>	263.1±14.3	314.0±14.4 <sup>*b,d</sup>	282.1±18.4	305.0±16.0 <sup>**e</sup>
Acetic acid (%)	75.3±0.55	73.4±1.01	75.0±0.55	74.23±1.28	75.2±0.46	73.7±0.75
Propionic acid (%)	16.4±0.74	15.9±0.70	17.3±0.63	17.0±0.94	15.3±0.55	15.0±0.71
Butyric acid (%)	8.25±0.44	10.73±0.56 <sup>*a</sup>	7.80±0.29	8.67±0.70 <sup>i,b,d</sup>	9.56±0.30	11.28±0.67 <sup>c</sup>

Data are expressed as mean ± SEM. Groups: CPF0inu0 n = 14, CPF0inu1 n = 12, CPF1inu0 n = 15, CPF1inu1 n = 11, CPF3.5inu0 n = 13, CPF3.5inu1 n = 14.

<sup>a</sup> CPF0inu0 vs. CPF0inu1

<sup>b</sup> CPF1inu0 vs. CPF1inu1

<sup>c</sup> CPF3.5inu0 vs. CPF3.5inu1

<sup>d</sup> CPF0inu1 vs. CPF1inu1

<sup>e</sup> CPF0inu1 vs. CPF3.5inu1

Signification

\*  $p < 0.05$

\*\*  $p < 0.01$ ; i = 0.058

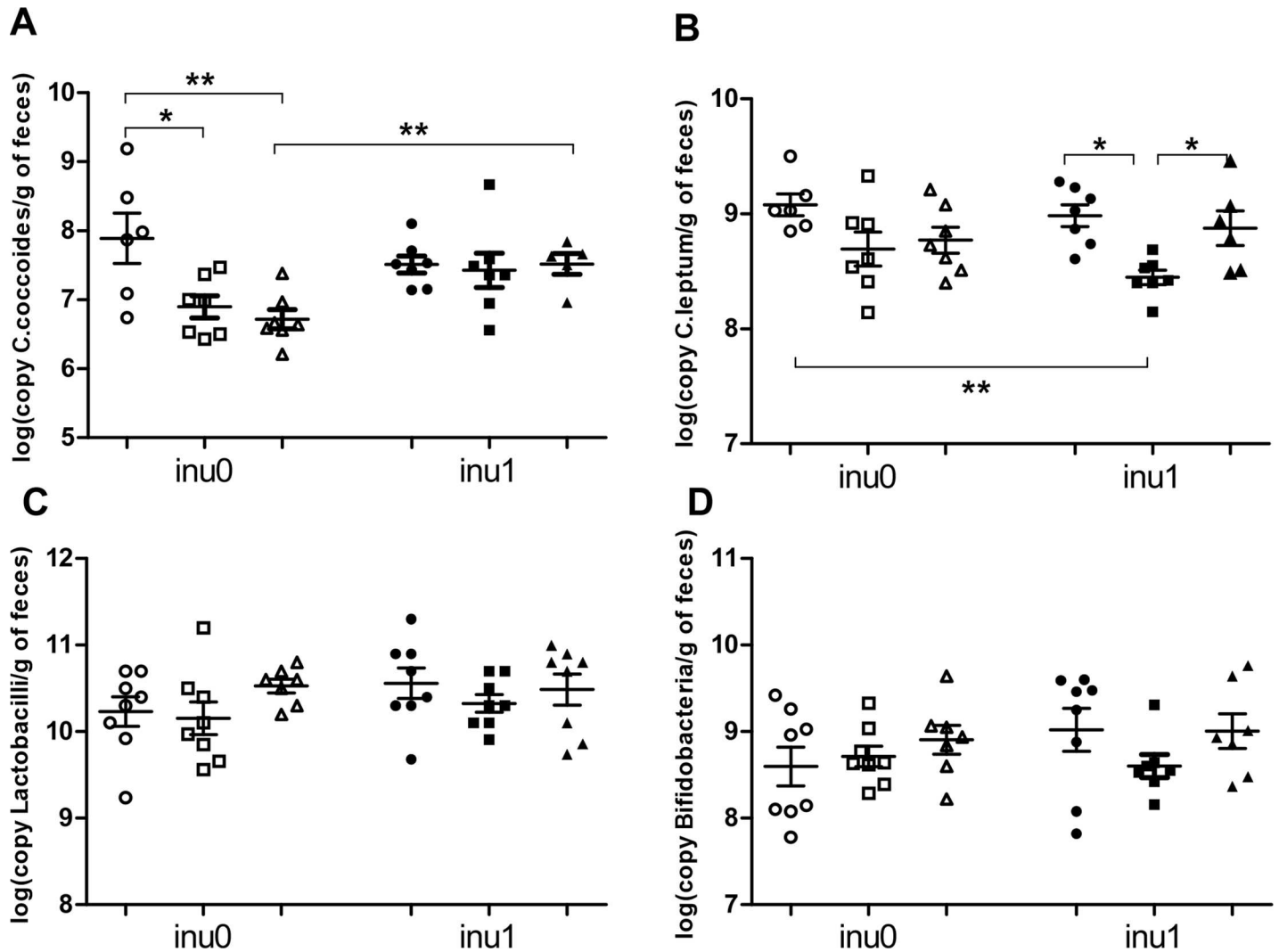
doi:10.1371/journal.pone.0164614.t002

*prausnitzii* or others clostridial clusters such as I, III, XI, XV, XVI [48] which use acetate produced by Bifidobacteria [49]. To our knowledge, we are the first to describe a decrease in *Firmicutes*, *C. coccoides* and *C. leptum* group in particular, after a perinatal CPF exposure in rats. This microbiota pattern resembles what we observed in the Human diabetic subjects with a decrease of *Firmicutes* and particularly butyrate-producing Clostridiales [50,51], *Bacteroides vulgates*, and Bifidobacteria [52]. It is well known that aberrant intestinal microbiota can induce a translocation of bacterial fragments and the development of “metabolic endotoxemia”, leading to systemic inflammation and insulin resistance [17]. Access to inulin may counteract endotoxemia in CPF-exposed rats as it alleviates the decrease in the phylum *Firmicutes* and clostridial clusters XIVa (*C. coccoides*) and IV (*C. leptum*).

In addition to the beneficial effect of inulin in CPF-exposed rats on microbiota and SCFA production, our results showed that it modified the metabolic status of the CPF-exposed rats according to the dose used. In CPF3.5 group, free access to inulin induced an increase in insulin, triglycerides and alanine transferase in the serum to a level close to those of control groups, showing that inulin restored some features of the disturbed metabolic profile in CPF3.5-exposed animals. Conversely, in CPF1-exposed rats, inulin decreased insulin secretion, initially high in CPF1-exposed rats, to a level comparable to those of control animals. Moreover, prebiotics increased the expression of IRβ in CPF1-exposed rats suggesting an enhancement of insulin sensitivity and a decrease of insulin resistance in these animals. Thus, altogether, our results show that our study is the first to demonstrate that bringing a continuous source of prebiotic to the mother and the infant, allowed young rats to alleviate the side effect of long-term exposure to CPF on their metabolism during their adult lives.

## Conclusions

In summary, the current results indicate that CPF exposure during pre- and postnatal period may be a limiting factor, at least in rodents, for the onset of a normal regulation of metabolism in the offspring during their adult life. It is suggested that organophosphate insecticides can increase the risk of diabetes mellitus. We propose that prebiotics, which have the ability to alter the microbiota in a positive manner, is a safe and cost-effective nutritional strategy to



**Fig 8. Bacterial genus profile.** (A) *C. coccoides* group; (B) *C. leptum* group; (C) Lactobacilli; (D) Bifidobacteria are expressed as log(number of copy/g of feces) mean  $\pm$  SEM [CPF0inu0 n = 6-8/CPF1inu0 n = 7-8/CPF3.5 n = 7/CPF0inu1 n = 7-8/CPF1inu1 n = 7-8/CPF3.5inu1 n = 5-8]. Control groups (CPF0, circles), CPF-exposed groups (CPF1: 1 mg/kg/day, squares; CPF3.5: CPF3.5 mg/kg/day, triangles) or inulin groups (black symbol). Significance \*p<0.05; \*\*p<0.01,\*\*\*p<0.001.

doi:10.1371/journal.pone.0164614.g008

counteract CPF insulin resistance and diabetes in later life. Human clinical trials should be undertaken to confirm these effects. However, additional basic research is necessary to better understand the crosstalk between microbiota and the host in order to elucidate the exact mechanism by which, microbiota alleviates the metabolic defects induced by CPF in adults. Experiments are in progress to determine whether the gut/brain axis is also involved in such regulation.

### Supporting Information

**S1 Table. Dam's body weight, food and drinking intake during gestation and lactation periods.** Data are expressed as mean  $\pm$  SEM and analyzed by Mann Withney test. Groups: CPF0inu0, CPF0inu1, CPF1inu0, CPF1inu1, CPF3.5inu0, CPF3.5inu1. (DOCX)



**S2 Table. Pup's food and drinking intake from PND 21 to PND 60.** Data are expressed as mean  $\pm$  SEM and analyzed by Mann Withney test. Groups: CPF0inu0, CPF0inu1, CPF1inu0, CPF1inu1, CPF3.5inu0, CPF3.5inu1. (DOCX)

## Acknowledgments

The authors thanks, Ludovic Didier, Virginie Jeanblanc and Anais Jovelet for technical assistance; Camille Mayeur for providing technical assistance and for relevant comments. The authors wish to thank Ask Scientific LTD, a company of proofreading services, for English editing.

## Author Contributions

**Conceptualization:** JR HKC LN.

**Data curation:** JR HKC LN.

**Formal analysis:** VB NBJ.

**Funding acquisition:** HKC FD VB LN.

**Investigation:** JR LL FD LR SD GE HKC LN.

**Methodology:** JR HKC LN.

**Project administration:** HKC LN.

**Resources:** HKC LL FD LR LN.

**Supervision:** LN.

**Validation:** HKC LN.

**Visualization:** SD GE LL NBJ LR FD VB HKC LN.

**Writing – original draft:** JR LN.

**Writing – review & editing:** JR HKC LN.

## References

1. Wild S, Roglic G, Green A, Sicree R, King H. Global prevalence of diabetes: estimates for the year 2000 and projections for 2030. *Diabetes Care*. 2004 May; 27(5):1047–53. PMID: [15111519](#)
2. Jaacks LM, Staimez LR. Association of persistent organic pollutants and non-persistent pesticides with diabetes and diabetes-related health outcomes in Asia: A systematic review. *Environ Int*. 2015 Mar; 76:57–70. doi: [10.1016/j.envint.2014.12.001](#) PMID: [25545233](#)
3. Saldana TM, Basso O, Hoppin JA, Baird DD, Knott C, Blair A, et al. Pesticide exposure and self-reported gestational diabetes mellitus in the Agricultural Health Study. *Diabetes Care*. 2007 Mar; 30(3):529–34. doi: [10.2337/dc06-1832](#) PMID: [17327316](#)
4. Taylor KW, Novak RF, Anderson HA, Birnbaum LS, Blystone C, Devito M, et al. Evaluation of the association between persistent organic pollutants (POPs) and diabetes in epidemiological studies: a national toxicology program workshop review. *Environ Health Perspect*. 2013 Jul; 121(7):774–83. doi: [10.1289/ehp.1205502](#) PMID: [23651634](#)
5. Thayer KA, Heindel JJ, Bucher JR, Gallo MA. Role of environmental chemicals in diabetes and obesity: a National Toxicology Program workshop review. *Environ Health Perspect*. 2012 Jun; 120(6):779–89. doi: [10.1289/ehp.1104597](#) PMID: [22296744](#)

6. Zhang Y, Han S, Liang D, Shi X, Wang F, Liu W, et al. Prenatal exposure to organophosphate pesticides and neurobehavioral development of neonates: a birth cohort study in Shenyang, China. *PLoS One*. 2014; 9(2):e88491. doi: [10.1371/journal.pone.0088491](https://doi.org/10.1371/journal.pone.0088491) PMID: [24551109](https://pubmed.ncbi.nlm.nih.gov/24551109/)
7. Saunders M, Magnanti BL, Correia Carreira S, Yang A, Alamo-Hernández U, Riojas-Rodriguez H, et al. Chlorpyrifos and neurodevelopmental effects: a literature review and expert elicitation on research and policy. *Environ Health Glob Access Sci Source*. 2012; 11 Suppl 1:S5.
8. Kwong TC. Organophosphate pesticides: biochemistry and clinical toxicology. *Ther Drug Monit*. 2002 Feb; 24(1):144–9. PMID: [11805735](https://pubmed.ncbi.nlm.nih.gov/11805735/)
9. Slotkin TA, Brown KK, Seidler FJ. Developmental exposure of rats to chlorpyrifos elicits sex-selective hyperlipidemia and hyperinsulinemia in adulthood. *Environ Health Perspect*. 2005 Oct; 113(10):1291–4. doi: [10.1289/ehp.8133](https://doi.org/10.1289/ehp.8133) PMID: [16203236](https://pubmed.ncbi.nlm.nih.gov/16203236/)
10. Slotkin TA. Does early-life exposure to organophosphate insecticides lead to prediabetes and obesity? *Reprod Toxicol Elmsford N*. 2011 Apr; 31(3):297–301.
11. Barker DJP, Lampl M. Commentary: The meaning of thrift. *Int J Epidemiol*. 2013 Oct; 42(5):1229–30. doi: [10.1093/ije/dyt131](https://doi.org/10.1093/ije/dyt131) PMID: [24159068](https://pubmed.ncbi.nlm.nih.gov/24159068/)
12. Khan MT, Nieuwdorp M, Bäckhed F. Microbial modulation of insulin sensitivity. *Cell Metab*. 2014 Nov 4; 20(5):753–60. doi: [10.1016/j.cmet.2014.07.006](https://doi.org/10.1016/j.cmet.2014.07.006) PMID: [25176147](https://pubmed.ncbi.nlm.nih.gov/25176147/)
13. Hur KY, Lee M-S. Gut Microbiota and Metabolic Disorders. *Diabetes Metab J*. 2015 Jun; 39(3):198–203. doi: [10.4093/dmj.2015.39.3.198](https://doi.org/10.4093/dmj.2015.39.3.198) PMID: [26124989](https://pubmed.ncbi.nlm.nih.gov/26124989/)
14. Tremaroli V, Bäckhed F. Functional interactions between the gut microbiota and host metabolism. *Nature*. 2012 Sep 13; 489(7415):242–9. doi: [10.1038/nature11552](https://doi.org/10.1038/nature11552) PMID: [22972297](https://pubmed.ncbi.nlm.nih.gov/22972297/)
15. Cani PD, Everard A, Duparc T. Gut microbiota, enteroendocrine functions and metabolism. *Curr Opin Pharmacol*. 2013 Dec; 13(6):935–40. doi: [10.1016/j.coph.2013.09.008](https://doi.org/10.1016/j.coph.2013.09.008) PMID: [24075718](https://pubmed.ncbi.nlm.nih.gov/24075718/)
16. Tomas J, Reygnier J, Mayeur C, Ducroc R, Bouet S, Bridonneau C, et al. Early colonizing *Escherichia coli* elicits remodeling of rat colonic epithelium shifting toward a new homeostatic state. *ISME J*. 2015 Jan; 9(1):46–58. doi: [10.1038/ismej.2014.111](https://doi.org/10.1038/ismej.2014.111) PMID: [25012905](https://pubmed.ncbi.nlm.nih.gov/25012905/)
17. Cani PD, Amar J, Iglesias MA, Poggi M, Knauf C, Bastelica D, et al. Metabolic endotoxemia initiates obesity and insulin resistance. *Diabetes*. 2007 Jul; 56(7):1761–72. doi: [10.2337/db06-1491](https://doi.org/10.2337/db06-1491) PMID: [17456850](https://pubmed.ncbi.nlm.nih.gov/17456850/)
18. Larsen N, Vogensen FK, van den Berg FWJ, Nielsen DS, Andreasen AS, Pedersen BK, et al. Gut microbiota in human adults with type 2 diabetes differs from non-diabetic adults. *PLoS One*. 2010; 5(2):e9085. doi: [10.1371/journal.pone.0009085](https://doi.org/10.1371/journal.pone.0009085) PMID: [20140211](https://pubmed.ncbi.nlm.nih.gov/20140211/)
19. Cani PD, Joly E, Horsmans Y, Delzenne NM. Oligofructose promotes satiety in healthy human: a pilot study. *Eur J Clin Nutr*. 2006 May; 60(5):567–72. doi: [10.1038/sj.ejcn.1602350](https://doi.org/10.1038/sj.ejcn.1602350) PMID: [16340949](https://pubmed.ncbi.nlm.nih.gov/16340949/)
20. Cani PD, Delzenne NM. The role of the gut microbiota in energy metabolism and metabolic disease. *Curr Pharm Des*. 2009; 15(13):1546–58. PMID: [19442172](https://pubmed.ncbi.nlm.nih.gov/19442172/)
21. Everard A, Lazarevic V, Derrien M, Girard M, Muccioli GG, Muccioli GM, et al. Responses of gut microbiota and glucose and lipid metabolism to prebiotics in genetic obese and diet-induced leptin-resistant mice. *Diabetes*. 2011 Nov; 60(11):2775–86. doi: [10.2337/db11-0227](https://doi.org/10.2337/db11-0227) PMID: [21933985](https://pubmed.ncbi.nlm.nih.gov/21933985/)
22. Byrne CS, Chambers ES, Morrison DJ, Frost G. The role of short chain fatty acids in appetite regulation and energy homeostasis. *Int J Obes* 2005. 2015 Sep; 39(9):1331–8.
23. Everard A, Cani PD. Diabetes, obesity and gut microbiota. *Best Pract Res Clin Gastroenterol*. 2013 Feb; 27(1):73–83. doi: [10.1016/j.bpg.2013.03.007](https://doi.org/10.1016/j.bpg.2013.03.007) PMID: [23768554](https://pubmed.ncbi.nlm.nih.gov/23768554/)
24. Geurts L, Neyrinck AM, Delzenne NM, Knauf C, Cani PD. Gut microbiota controls adipose tissue expansion, gut barrier and glucose metabolism: novel insights into molecular targets and interventions using prebiotics. *Benef Microbes*. 2014 Mar; 5(1):3–17. doi: [10.3920/BM2012.0065](https://doi.org/10.3920/BM2012.0065) PMID: [23886976](https://pubmed.ncbi.nlm.nih.gov/23886976/)
25. Joly Condette C, Bach V, Mayeur C, Gay-Quéheillard J, Khorsi-Cauet H. Chlorpyrifos Exposure During Perinatal Period Affects Intestinal Microbiota Associated With Delay of Maturation of Digestive Tract in Rats. *J Pediatr Gastroenterol Nutr*. 2015 Jul; 61(1):30–40. doi: [10.1097/MPG.0000000000000734](https://doi.org/10.1097/MPG.0000000000000734) PMID: [25643018](https://pubmed.ncbi.nlm.nih.gov/25643018/)
26. Elmhiri G, Barella LF, Vieau D, Camous S, Mathias PCF, Abdennebi-Najar L. Acute exposure to a precursor of advanced glycation end products induces a dual effect on the rat pancreatic islet function. *Int J Endocrinol*. 2014; 2014:378284. doi: [10.1155/2014/378284](https://doi.org/10.1155/2014/378284) PMID: [25484898](https://pubmed.ncbi.nlm.nih.gov/25484898/)
27. Abdennebi-Najar L, Desai M, Han G, Casillas E, Jean D, Arie G, et al. Basal, endogenous leptin is metabolically active in newborn rat pups. *J Matern-Fetal Neonatal Med Off J Eur Assoc Perinat Med Fed Asia Ocean Perinat Soc Int Soc Perinat Obstet*. 2011 Dec; 24(12):1486–91.
28. Mayeur C, Gratadoux J-J, Bridonneau C, Chegdani F, Larroque B, Kapel N, et al. Faecal D/L lactate ratio is a metabolic signature of microbiota imbalance in patients with short bowel syndrome. *PLoS One*. 2013; 8(1):e54335. doi: [10.1371/journal.pone.0054335](https://doi.org/10.1371/journal.pone.0054335) PMID: [23372709](https://pubmed.ncbi.nlm.nih.gov/23372709/)

29. Etreiki C, Gadonna-Widehem P, Mangin I, Coëffier M, Delayre-Orthez C, Anton PM. Juvenile ferric iron prevents microbiota dysbiosis and colitis in adult rodents. *World J Gastroenterol*. 2012 Jun 7; 18(21):2619–29. doi: [10.3748/wjg.v18.i21.2619](https://doi.org/10.3748/wjg.v18.i21.2619) PMID: [22690070](https://pubmed.ncbi.nlm.nih.gov/22690070/)
30. Lecerf J- M, Dépeint F, Clerc E, Dugenet Y, Niamba CN, Rhazi L, et al. Xylo-oligosaccharide (XOS) in combination with inulin modulates both the intestinal environment and immune status in healthy subjects, while XOS alone only shows prebiotic properties. *Br J Nutr*. 2012 Nov 28; 108(10):1847–58. doi: [10.1017/S0007114511007252](https://doi.org/10.1017/S0007114511007252) PMID: [22264499](https://pubmed.ncbi.nlm.nih.gov/22264499/)
31. Firmin S, Bahi-Jaber N, Abdennebi-Najar L. Food contaminants and programming of type 2 diabetes: recent findings from animal studies. *J Dev Orig Health Dis*. 2016 Jun 13;1–8.
32. Barker DJP. The developmental origins of insulin resistance. *Horm Res*. 2005; 64 Suppl 3:2–7. doi: [10.1159/000089311](https://doi.org/10.1159/000089311) PMID: [16439838](https://pubmed.ncbi.nlm.nih.gov/16439838/)
33. Lassiter TL, Ryde IT, Mackillop EA, Brown KK, Levin ED, Seidler FJ, et al. Exposure of neonatal rats to parathion elicits sex-selective reprogramming of metabolism and alters the response to a high-fat diet in adulthood. *Environ Health Perspect*. 2008 Nov; 116(11):1456–62. doi: [10.1289/ehp.11673](https://doi.org/10.1289/ehp.11673) PMID: [19057696](https://pubmed.ncbi.nlm.nih.gov/19057696/)
34. Lassiter TL, Barone S, Padilla S. Ontogenetic differences in the regional and cellular acetylcholinesterase and butyrylcholinesterase activity in the rat brain. *Brain Res Dev Brain Res*. 1998 Jan 14; 105(1):109–23.
35. Mansour SA, Mossa A- TH. Adverse effects of exposure to low doses of chlorpyrifos in lactating rats. *Toxicol Ind Health*. 2011 Apr; 27(3):213–24. doi: [10.1177/0748233710384054](https://doi.org/10.1177/0748233710384054) PMID: [20870695](https://pubmed.ncbi.nlm.nih.gov/20870695/)
36. Saltiel AR, Kahn CR. Insulin signalling and the regulation of glucose and lipid metabolism. *Nature*. 2001 Dec 13; 414(6865):799–806. doi: [10.1038/414799a](https://doi.org/10.1038/414799a) PMID: [11742412](https://pubmed.ncbi.nlm.nih.gov/11742412/)
37. Meggs WJ, Brewer KL. Weight gain associated with chronic exposure to chlorpyrifos in rats. *J Med Toxicol Off J Am Coll Med Toxicol*. 2007 Sep; 3(3):89–93.
38. Tanvir EM, Afroz R, Chowdhury M, Gan SH, Karim N, Islam MN, et al. A model of chlorpyrifos distribution and its biochemical effects on the liver and kidneys of rats. *Hum Exp Toxicol*. 2015 Oct 30;
39. Joly Condet C, Khorsi-Cauet H, Morlière P, Zabijak L, Reygnier J, Bach V, et al. Increased gut permeability and bacterial translocation after chronic chlorpyrifos exposure in rats. *PLoS One*. 2014; 9(7): e102217. doi: [10.1371/journal.pone.0102217](https://doi.org/10.1371/journal.pone.0102217) PMID: [25019507](https://pubmed.ncbi.nlm.nih.gov/25019507/)
40. Michael MD, Kulkarni RN, Postic C, Previs SF, Shulman GI, Magnuson MA, et al. Loss of insulin signaling in hepatocytes leads to severe insulin resistance and progressive hepatic dysfunction. *Mol Cell*. 2000 Jul; 6(1):87–97. PMID: [10949030](https://pubmed.ncbi.nlm.nih.gov/10949030/)
41. Roberfroid M, Gibson GR, Hoyles L, McCartney AL, Rastall R, Rowland I, et al. Prebiotic effects: metabolic and health benefits. *Br J Nutr*. 2010 Aug; 104 Suppl 2:S1–63. doi: [10.1017/S0007114510003363](https://doi.org/10.1017/S0007114510003363) PMID: [20920376](https://pubmed.ncbi.nlm.nih.gov/20920376/)
42. Van de Wiele T, Boon N, Possemiers S, Jacobs H, Verstraete W. Inulin-type fructans of longer degree of polymerization exert more pronounced in vitro prebiotic effects. *J Appl Microbiol*. 2007 Feb; 102(2):452–60. doi: [10.1111/j.1365-2672.2006.03084.x](https://doi.org/10.1111/j.1365-2672.2006.03084.x) PMID: [17241351](https://pubmed.ncbi.nlm.nih.gov/17241351/)
43. Scott KP, Martin JC, Duncan SH, Flint HJ. Prebiotic stimulation of human colonic butyrate-producing bacteria and bifidobacteria, in vitro. *FEMS Microbiol Ecol*. 2014 Jan; 87(1):30–40. doi: [10.1111/1574-6941.12186](https://doi.org/10.1111/1574-6941.12186) PMID: [23909466](https://pubmed.ncbi.nlm.nih.gov/23909466/)
44. Ramirez-Farias C, Slezak K, Fuller Z, Duncan A, Holtrop G, Louis P. Effect of inulin on the human gut microbiota: stimulation of *Bifidobacterium adolescentis* and *Faecalibacterium prausnitzii*. *Br J Nutr*. 2009 Feb; 101(4):541–50. doi: [10.1017/S0007114508019880](https://doi.org/10.1017/S0007114508019880) PMID: [18590586](https://pubmed.ncbi.nlm.nih.gov/18590586/)
45. Campbell JM, Fahey GC, Wolf BW. Selected indigestible oligosaccharides affect large bowel mass, cecal and fecal short-chain fatty acids, pH and microflora in rats. *J Nutr*. 1997 Jan; 127(1):130–6. PMID: [9040556](https://pubmed.ncbi.nlm.nih.gov/9040556/)
46. Weitkunat K, Schumann S, Petzke KJ, Blaut M, Loh G, Klaus S. Effects of dietary inulin on bacterial growth, short-chain fatty acid production and hepatic lipid metabolism in gnotobiotic mice. *J Nutr Biochem*. 2015 Sep; 26(9):929–37. doi: [10.1016/j.jnutbio.2015.03.010](https://doi.org/10.1016/j.jnutbio.2015.03.010) PMID: [26033744](https://pubmed.ncbi.nlm.nih.gov/26033744/)
47. Kimura I, Ozawa K, Inoue D, Imamura T, Kimura K, Maeda T, et al. The gut microbiota suppresses insulin-mediated fat accumulation via the short-chain fatty acid receptor GPR43. *Nat Commun*. 2013; 4:1829. doi: [10.1038/ncomms2852](https://doi.org/10.1038/ncomms2852) PMID: [23652017](https://pubmed.ncbi.nlm.nih.gov/23652017/)
48. Louis P, Scott KP, Duncan SH, Flint HJ. Understanding the effects of diet on bacterial metabolism in the large intestine. *J Appl Microbiol*. 2007 May; 102(5):1197–208. doi: [10.1111/j.1365-2672.2007.03322.x](https://doi.org/10.1111/j.1365-2672.2007.03322.x) PMID: [17448155](https://pubmed.ncbi.nlm.nih.gov/17448155/)
49. Duncan SH, Holtrop G, Lobley GE, Calder AG, Stewart CS, Flint HJ. Contribution of acetate to butyrate formation by human faecal bacteria. *Br J Nutr*. 2004 Jun; 91(6):915–23. doi: [10.1079/BJN20041150](https://doi.org/10.1079/BJN20041150) PMID: [15182395](https://pubmed.ncbi.nlm.nih.gov/15182395/)

50. Karlsson FH, Tremaroli V, Nookaew I, Bergström G, Behre CJ, Fagerberg B, et al. Gut metagenome in European women with normal, impaired and diabetic glucose control. *Nature*. 2013 Jun 6; 498(7452):99–103. doi: [10.1038/nature12198](https://doi.org/10.1038/nature12198) PMID: [23719380](https://pubmed.ncbi.nlm.nih.gov/23719380/)
51. Qin J, Li Y, Cai Z, Li S, Zhu J, Zhang F, et al. A metagenome-wide association study of gut microbiota in type 2 diabetes. *Nature*. 2012 Oct 4; 490(7418):55–60. doi: [10.1038/nature11450](https://doi.org/10.1038/nature11450) PMID: [23023125](https://pubmed.ncbi.nlm.nih.gov/23023125/)
52. Wu X, Ma C, Han L, Nawaz M, Gao F, Zhang X, et al. Molecular characterisation of the faecal microbiota in patients with type II diabetes. *Curr Microbiol*. 2010 Jul; 61(1):69–78. doi: [10.1007/s00284-010-9582-9](https://doi.org/10.1007/s00284-010-9582-9) PMID: [20087741](https://pubmed.ncbi.nlm.nih.gov/20087741/)